

## Variation in Selectivity of Univalent Cations in Slime Mold *Physarum Polycephalum* Caused by Reception of Polyvalent Cations

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*Summary.* Specificity of reception on 1:1 electrolytes in the slime mold *Physarum polycephalum* was investigated in the presence of polyvalent cations in media. Membrane potential and motive force of tactic movement were examined with the aid of the double chamber method, and the zeta potential at the membrane surface of the slime mold was measured by electrophoretic mobility. The results obtained are summarized as follows: (1) The presence of polyvalent cations (e.g.,  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Sr}^{2+}$ ,  $\text{Ba}^{2+}$ ,  $\text{La}^{3+}$ ,  $\text{Th}^{4+}$ ) in medium led to an increase in threshold concentration,  $C_{th}$ , determined from the potential measurements for Na- or Li-salts, and to a decrease in  $C_{th}$  for K-, Rb- or  $\text{NH}_4$ -salts.  $C_{th}$  for 1:1 electrolytes changed discontinuously when the concentration of polyvalent cations in medium exceeded their respective thresholds. (2) The  $C_{th}$  determined from chemotaxis agreed with that from the potential response both in the presence and absence of polyvalent cations. (3) Sequence of selectivity of univalent cations varied extensively in the presence of polyvalent cations. (4) Changes in the zeta potential induced by NaCl reception agreed with those in the membrane potential even in the presence of  $\text{Ca}^{2+}$  in medium. (5) The  $C_{th}$  for reception of NaCl changed sharply at about 12°C in the presence of polyvalent cations, while that for KCl was independent of the temperature.

Conformational changes in surface membrane of the slime mold in response to reception of polyvalent cations were then discussed in relation to the discrimination of univalent cations.

It is well known that divalent cations interact with univalent cations antagonistically in many biological membrane systems. For example, the responses to univalent cations are suppressed by the presence of divalent cations in a fish lateral line organ [15], and in frog taste organs [14, 17]. The abrupt depolarizations in cell potential in squid giant axons and in the excitable protoplasmic droplets isolated from *Nitella* caused by univalent cations are suppressed by  $\text{Ca}^{2+}$  added in media [11, 16, 19, 20]. The antagonistic interaction between univalent and divalent cations

is considered to play an indispensable role in the appearance of various functions in biological membranes, but the relevant physicochemical mechanism remains almost unknown at the present.

The plasmodium of the true slime mold *Physarum polycephalum* exhibits positive or negative chemotaxis to various kinds of chemical substances [1, 3, 9, 18, 21, 22, 23]. The plasmodium of the slime mold is a suitable experimental organism to study the physicochemical mechanism of chemoreception and taxis at the receptor membrane level, because the plasmodium is a large aggregate of protoplasm without boundary membrane separating the cellular components. In a previous paper [23], we demonstrated that the motive force of chemotactic movement and the membrane potential of the slime mold changed simultaneously at a certain threshold concentration,  $C_{th}$ , for respective chemicals, and that the slime mold discriminated various univalent cations. The sequence or the selectivity for univalent cations determined from the  $C_{th}$  values was  $\text{Li}^+ < \text{K}^+ < \text{Na}^+ < \text{Rb}^+ < \text{Cs}^+ < \text{NH}_4^+$  in pure water. In the present study, we investigated the selectivity of 1:1 electrolytes reception of the slime mold in the presence of polyvalent cations in the external solution, and we examined variations of univalent cation selectivity accompanying the reception of polyvalent cations. The sequence of cation selectivity described above will be shown to change discontinuously when  $\text{Ca}^{2+}$  or other polyvalent cations exceeded their respective threshold concentrations. The implication of these results is discussed in relation to the structural change of the surface membrane of the slime mold.

## Materials and Methods

The true slime mold *Physarum polycephalum* used in the present study was kindly furnished by Prof. N. Kamiya of Osaka University, and cultured by the method employed by Camp [2]. Microplasmodia of the slime mold *Physarum polycephalum* used in the electrophoretic study were a gift of Prof. J. Ohta at Ochanomizu University. Culture of the microplasmodia was performed according to the method proposed by Daniel and Rusch [5].

Measurements of the membrane potential and the motive force of protoplasmic movement were performed by using a double chamber method proposed by Kamiya [12, 13], the details of which were described in the previous paper [23]. Two pieces of plasmodia were placed in two compartments which were connected by a plasmodial strand through a narrow ditch. The opening of the ditch was filled with silicone grease and the chambers were covered by a slide glass so as to make the two compartments airtight and to insulate them electrically, except for the plasmodial strand. The basis of the method lies in the fact that the protoplasmic streaming can be controlled by applying a difference in hydrostatic pressure between the two compartments, and that the potential difference between them affords a measure of the membrane potential [21, 22, 23]. The

effect of the presence of polyvalent cations in medium on the response to 1:1 electrolytes was examined as follows: aqueous solution of a given concentration of a polyvalent cation was placed in both compartments, and then the solution in one compartment was replaced with a 1:1 electrolyte solution containing the same concentration of the polyvalent cation. The differences in the membrane potential and motive force of taxis were measured successively by increasing the added 1:1 salt concentration.

The zeta potential of the microplasmidia of the slime mold was measured by the method accomplished by Hato *et al.* [10], by using a microelectrophoretic apparatus (Carl Zeiss, Cytopherometer). In the present study, a solution containing 10 mM sucrose and 1 mM NaCl was used as the standard solution. The zeta potentials were measured by increasing concentration of 1:1 electrolyte with a given concentration of a polyvalent cation added to the standard solution.

All experiments were performed at a room temperature,  $20 \pm 0.5^\circ\text{C}$ , except the experiments studying the temperature effect. Dependency of the threshold on the temperature was examined by regulating the room temperature within  $\pm 0.5^\circ\text{C}$  at a desired value ranging between 6 and  $30^\circ\text{C}$ . All chemicals used were analytical grade without further purification. Water used as solvent was distilled twice by glass vessels.

## Results

### *Effects of $\text{Ca}^{2+}$ on Reception of 1:1 Electrolytes*

Fig. 1 shows the dependences of the membrane potential,  $\Delta\phi$ , zeta potential,  $\zeta$ , and of chemotactic motive force,  $\overline{\Delta P}$ , on NaCl concentration. In Fig. 1,  $\Delta\zeta$  is defined by  $\Delta\zeta = \zeta - \zeta_0$ , where  $\zeta_0$  stands for the zeta potential in NaCl solution whose concentration is lower than the threshold. The value of  $\zeta_0$  depends slightly on the  $\text{CaCl}_2$  concentration when it exceeds  $6 \times 10^{-5} \text{ M}$ , e.g.,  $\zeta_0 = -48 \text{ mV}$  in the standard solution and in  $2 \times 10^{-5} \text{ M}$ , while  $\zeta_0 = -45 \text{ mV}$  in  $10^{-4} \text{ M CaCl}_2$ . In the absence of  $\text{CaCl}_2$  in medium,  $\Delta\phi$  started to change in the direction of depolarization at the concentration of  $2.5 \times 10^{-3} \text{ M NaCl}$  as illustrated by  $\circ$  marks. The variation of the zeta potential,  $\Delta\zeta$  ( $\bullet$  marks), agreed with that of  $\Delta\phi$ . The plasmodium showed a negative taxis at the same concentration and the motive force of tactic movement  $\overline{\Delta P}$  stayed at the level of about 10 cm  $\text{H}_2\text{O}$  with further increase of NaCl concentration. The negative value of  $\overline{\Delta P}$  represents a negative taxis. Note that the presence of  $2 \times 10^{-5} \text{ M CaCl}_2$  in medium led to appreciable influence on the  $\Delta\phi$ ,  $\Delta\zeta$ , and  $\overline{\Delta P}$  vs.  $\log[\text{NaCl}]$  relations. In the presence of  $10^{-4} \text{ M CaCl}_2$  in medium, however,  $\Delta\phi$  and  $\Delta\zeta$  started to change at the concentration of  $10^{-2} \text{ M NaCl}$ , and the plasmodium showed negative taxis at the same concentration ( $\bullet$  marks). In other words, the presence of  $10^{-4} \text{ M CaCl}_2$  in medium increased the threshold concentration,  $C_{th}$ , for NaCl from  $2.5 \times 10^{-3}$  to  $10^{-2} \text{ M}$ . The data shown in Fig. 1 illustrate that the changes in  $\Delta\phi$  in response to NaCl agree with the corresponding

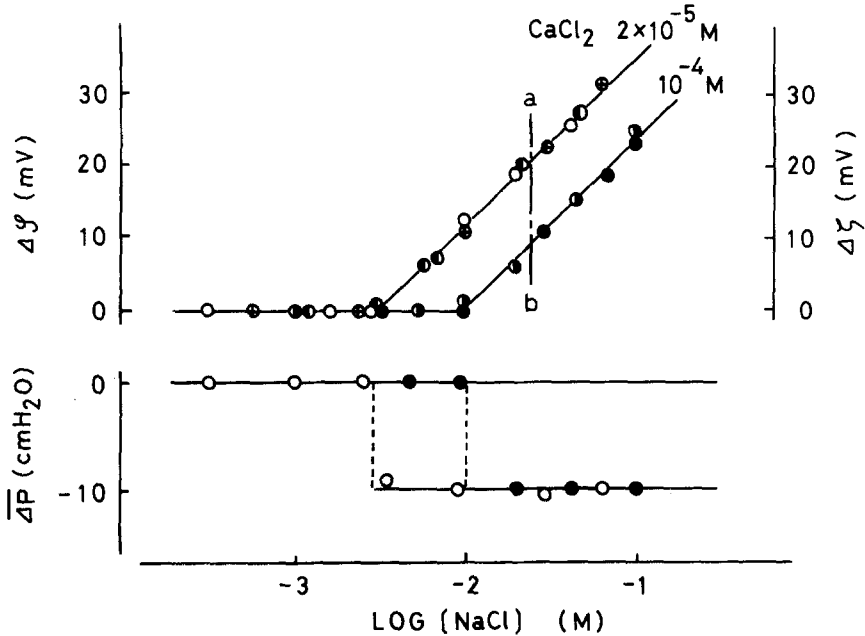


Fig. 1. Changes in membrane potential,  $\Delta\phi$ , zeta potential,  $\Delta\zeta = (\zeta - \zeta_0)$ , and motive force of taxis,  $\overline{\Delta P}$ , as a function of NaCl concentration in the presence and absence of  $\text{CaCl}_2$ .  $\circ$ :  $\Delta\phi$ ,  $\overline{\Delta P}$  in the absence of  $\text{CaCl}_2$  in media;  $\odot$ :  $\Delta\phi$  in the presence of  $2 \times 10^{-5} \text{ M}$   $\text{CaCl}_2$ ;  $\bullet$ :  $\Delta\zeta$  in the presence of  $2 \times 10^{-5} \text{ M}$   $\text{CaCl}_2$ ;  $\ominus$ :  $\Delta\phi$ ,  $\overline{\Delta P}$  in the presence of  $10^{-4} \text{ M}$   $\text{CaCl}_2$ ;  $\ominus$ :  $\Delta\zeta$  in the presence of  $10^{-4} \text{ M}$   $\text{CaCl}_2$ . Depolarization of the membrane potential is taken as positive

changes in zeta potential in the absence and presence of  $\text{CaCl}_2$ . The agreement between zeta and membrane potentials was obtained in all concentrations of  $\text{CaCl}_2$  examined, although a discontinuous change in the  $C_{th}$  for NaCl reception resulted at a certain concentration of  $\text{CaCl}_2$ . The implication of these results will be discussed later.

Fig. 2 shows a similar plot of  $\Delta\phi$  and  $\overline{\Delta P}$  as a function of KCl concentration in the absence and the presence of  $3 \times 10^{-4} \text{ M}$   $\text{CaCl}_2$  in the external solution. In the absence of  $\text{CaCl}_2$ , the variation of  $\Delta\phi$  took place at  $10^{-3} \text{ M}$  KCl, above which point the slime mold showed negative chemotaxis. Contrary to NaCl, the presence of  $3 \times 10^{-4} \text{ M}$   $\text{CaCl}_2$  led to a marked decrease in the  $C_{th}$  for KCl reception where a depolarization of  $\Delta\phi$  took place at about  $10^{-4} \text{ M}$  KCl, and  $\Delta\phi$  changed in two steps, in the manner seen in Fig. 2. The slime mold showed a weak positive taxis (maximum of  $\overline{\Delta P} \cong 5 \text{ cm H}_2\text{O}$ ) in the range of  $10^{-4} - 10^{-3} \text{ M}$  KCl concentration.  $\overline{\Delta P}$  became zero at  $10^{-3} \text{ M}$  KCl, and the slime mold showed negative taxis when the concentration of KCl increased further. Since the

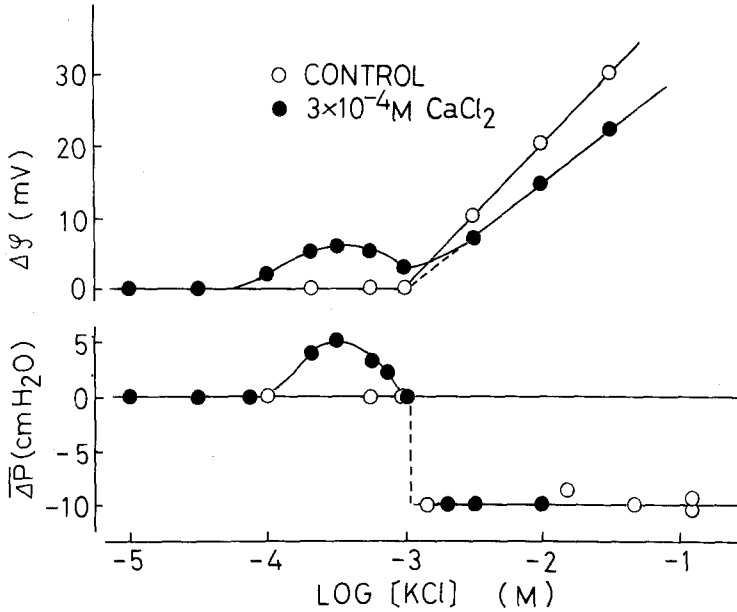


Fig. 2. Changes in membrane potential,  $\Delta\phi$ , and in chemotactic motive force,  $\overline{\Delta P}$ , as a function of KCl concentration in the presence of  $3 \times 10^{-4}$  M  $\text{CaCl}_2$  (●) and in the absence of  $\text{CaCl}_2$  (○)

slime mold showed appreciable changes both in  $\Delta\phi$  and  $\overline{\Delta P}$  at  $10^{-4}$  M KCl, this point is referred to as the threshold for KCl in the presence of  $3 \times 10^{-4}$  M  $\text{CaCl}_2$ . Response to  $\text{NH}_4\text{Cl}$  or  $\text{RbCl}$  in the presence of  $\text{CaCl}_2$  was essentially in a fashion similar to that of KCl, i.e., an appreciable decrease in  $C_{th}$  with a depolarization in membrane potential and a weak positive taxis. On the other hand, responses of the slime mold to  $\text{LiCl}$  were similar to those for  $\text{NaCl}$  when  $\text{CaCl}_2$  concentration in medium exceeded  $6 \times 10^{-5}$  M, i.e., the threshold concentration of  $\text{CaCl}_2$  itself.

Fig. 3 summarizes the change in the  $C_{th}$  for chloride salts of Li, K, Na, Rb and  $\text{NH}_4$  as a function of  $\text{CaCl}_2$  concentration in media both in logarithmic scales. When the  $\text{CaCl}_2$  concentration exceeded the threshold for  $\text{CaCl}_2$ , the  $C_{th}$  for each chloride salt changed discontinuously. Fig. 3 shows that the  $C_{th}$  for  $\text{NaCl}$  and  $\text{LiCl}$  increased about 4 and 10 times, respectively, in the presence of  $\text{CaCl}_2$  above  $6 \times 10^{-5}$  M, as compared with the cases with no  $\text{CaCl}_2$  in media, i.e.,  $3 \times 10^{-3}$  M for  $\text{NaCl}$ , and  $5 \times 10^{-4}$  M for  $\text{LiCl}$ . When the concentration of  $\text{CaCl}_2$  exceeded  $10^{-3}$  M in the case of  $\text{NaCl}$ , and  $3 \times 10^{-3}$  M in the case of  $\text{LiCl}$ , the  $C_{th}$  for these salts returned to their respective original levels. Contrary to  $\text{NaCl}$  or  $\text{LiCl}$ , the  $C_{th}$  for  $\text{KCl}$ ,  $\text{RbCl}$  and  $\text{NH}_4\text{Cl}$  decreased

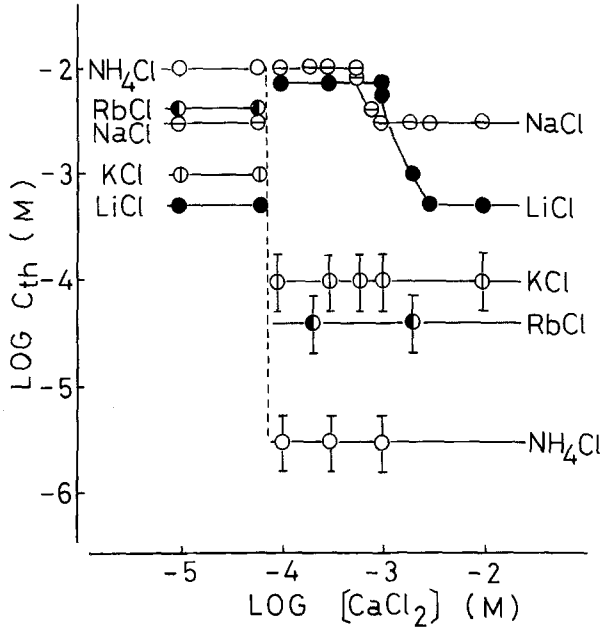


Fig. 3. The threshold concentrations,  $C_{th}$ , of LiCl, KCl, NaCl, RbCl and  $\text{NH}_4\text{Cl}$  as a function of concentration of  $\text{CaCl}_2$  added in the external solution

discontinuously and stayed at a constant level of  $10^{-4}$  M,  $4 \times 10^{-5}$  M and  $3 \times 10^{-6}$  M, respectively, with further increase of  $\text{CaCl}_2$  concentration. The slime mold remarkably changed the selectivity to univalent cations when  $\text{CaCl}_2$  concentration exceeded the threshold of  $\text{CaCl}_2$ , i.e.,  $6 \times 10^{-5}$  M. The sequence of univalent cation selectivity in the absence of  $\text{Ca}^{2+}$  ( $\text{Li}^+ < \text{K}^+ < \text{Na}^+ < \text{Rb}^+ < \text{NH}_4^+$ ) was changed to  $\text{NH}_4^+ < \text{Rb}^+ < \text{K}^+ < \text{Li}^+ < \text{Na}^+$ . When the concentration of  $\text{Ca}^{2+}$  in medium exceeded about  $10^{-3}$  M, the sequence became  $\text{NH}_4^+ < \text{Rb}^+ < \text{K}^+ < \text{Li}^+ < \text{Na}^+$ .

#### *Discrimination of Anions in 1:1 Electrolytes in the Presence of $\text{Ca}^{2+}$*

In the previous paper [23], we showed that the  $C_{th}$  for various 1:1 salts carrying a common cation with different anions fell on a straight line when  $\log C_{th}$  is plotted against the lyotropic number of anions,  $N$ . The lyotropic number of anions was determined from the relative concentration required to cause the coagulation of agar or protein solution [20, 24]. Fig. 4 shows the relation between  $\log C_{th}$  and  $N$  for NaCl, LiCl, KCl and  $\text{NH}_4\text{Cl}$  in the presence of  $3 \times 10^{-4}$  M  $\text{CaCl}_2$  and

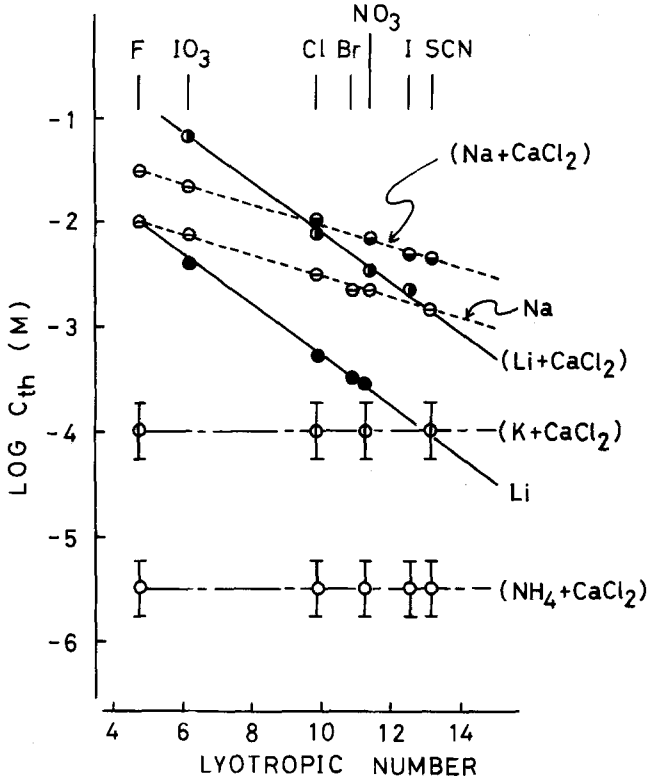


Fig. 4. Relations between the  $C_{th}$  for 1:1 type salts and the lyotropic number of anions in the presence of  $3 \times 10^{-4}$  M  $\text{CaCl}_2$  and in the absence of  $\text{CaCl}_2$ . Anion species are indicated at the top in the figure, and cations used are noted at the right of respective lines

with no  $\text{CaCl}_2$  in media. It is noted that the  $C_{th}$  of Na-salts with different anions increased the  $C_{th}$  by the same amount in the presence of  $\text{CaCl}_2$  compared with those in the absence of  $\text{CaCl}_2$ ; namely, the straight line in  $\log C_{th}$  vs.  $N$  plots for Na-salts in the presence of  $\text{CaCl}_2$  was shifted in parallel from the straight line for Na-salts without  $\text{CaCl}_2$ . The similar relation between  $\log C_{th}$  vs.  $N$  relation was observed for Li-salts as illustrated in Fig. 4. On the other hand, the  $C_{th}$  for K-salts decreased remarkably in the presence of  $3 \times 10^{-4}$  M  $\text{CaCl}_2$ , and stayed at a constant level,  $10^{-4}$  M, irrespective of the species of anions involved.  $\text{NH}_4$ -salts with different anions showed the similar dependency of  $C_{th}$  on  $N$  as shown in the figure.  $\text{NH}_4$ -salts with different anions showed the same value of  $C_{th}$ ,  $10^{-2}$  M, in pure water independent of anion species [23], which was changed to  $3 \times 10^{-6}$  M in the presence of  $\text{CaCl}_2$ .

As shown above, the  $C_{th}$  for receptions to 1:1 electrolytes of the slime mold in the presence of  $\text{CaCl}_2$  differed from one salt to the other

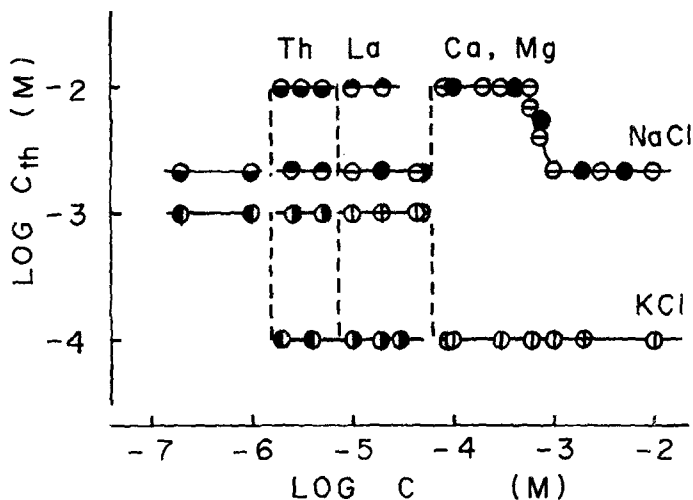


Fig. 5. The  $C_{th}$  of NaCl and KCl as a function of concentration of polyvalent cations added in media.  $\ominus$ :  $\text{Th}^{4+}$ ,  $\bullet$ :  $\text{La}^{3+}$ ,  $\circ$ :  $\text{Ca}^{2+}$ ,  $\bullet$ :  $\text{Mg}^{2+}$  for NaCl;  $\ominus$ :  $\text{Th}^{4+}$ ,  $\bullet$ :  $\text{La}^{3+}$ ,  $\circ$ :  $\text{Ca}^{2+}$ ,  $\ominus$ :  $\text{Mg}^{2+}$  for KCl

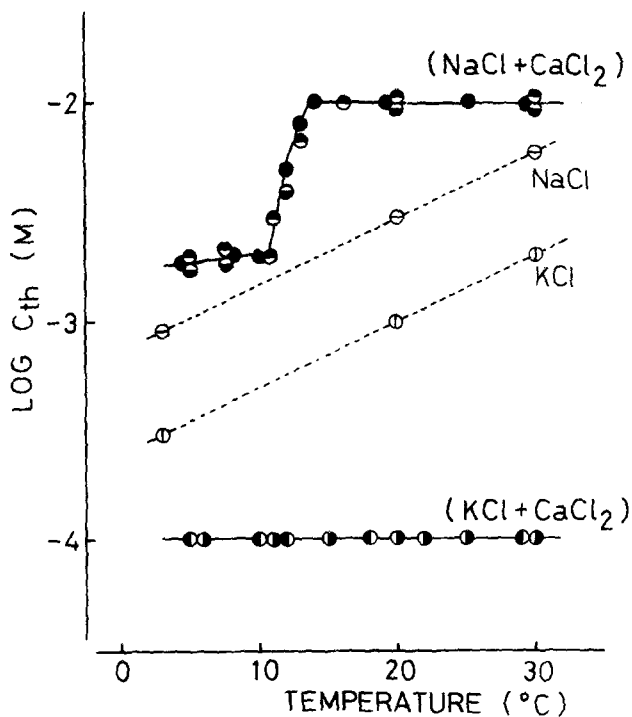


Fig. 6. Temperature dependences of the  $C_{th}$  for NaCl and KCl in the presence and absence of  $\text{CaCl}_2$ .  $\text{CaCl}_2$  concentration are  $10^{-4} \text{ M}$  ( $\bullet$ ,  $\circ$ ),  $3 \times 10^{-4} \text{ M}$  ( $\bullet$ ,  $\ominus$ ),  $6 \times 10^{-4} \text{ M}$  ( $\bullet$ ,  $\circ$ ), and  $0 \text{ M}$  ( $\ominus$ ,  $\circ$ ). Salt species in each cases are indicated in the figure at the right of respective lines



depending on the species of cation involved, and which were quite different from those in the absence of  $\text{Ca}^{2+}$ . Similar results were observed not only with  $\text{Ca}^{2+}$  but also with the other polyvalent cations.  $\text{Mg}^{2+}$ ,  $\text{Sr}^{2+}$ , and  $\text{Ba}^{2+}$  changed the selectivity for univalent cations discontinuously when the concentration exceeded  $6 \times 10^{-5} \text{ M}$  as in the case of  $\text{Ca}^{2+}$ .  $\text{La}^{3+}$  and  $\text{Th}^{4+}$  changed the selectivity for univalent cations at  $7 \times 10^{-6} \text{ M}$  and  $1.5 \times 10^{-6} \text{ M}$ , respectively. The discontinuous changes in the  $C_{th}$  for KCl and NaCl in the presence of various polyvalent cations are illustrated in Fig. 5 as a function of polyvalent cation in media. Note that the concentration of polyvalent cations which induced the discontinuous changes in  $C_{th}$  for univalent cation reception agreed with the threshold concentration of respective cations [23].

#### *Effects of Temperature on the $C_{th}$ for NaCl and KCl*

Fig. 6 shows the dependences of the  $C_{th}$  for NaCl and KCl on temperature in the absence and the presence of  $\text{CaCl}_2$ . The  $C_{th}$  for NaCl changed sharply at about  $12^\circ\text{C}$  when the concentration of  $\text{CaCl}_2$  exceeded its threshold,  $6 \times 10^{-5} \text{ M}$ . Contrary to the  $C_{th}$  for NaCl, the  $C_{th}$  for KCl did not show any temperature dependency and stayed at a constant level of  $10^{-4} \text{ M}$  when the concentration of  $\text{CaCl}_2$  in media exceeded  $6 \times 10^{-5} \text{ M}$ . These temperature dependencies of the  $C_{th}$  for KCl and NaCl in the presence of  $\text{CaCl}_2$  differ from those in the absence of  $\text{CaCl}_2$ , which are shown by broken lines in the figure. Similar results are obtained with polyvalent cations other than  $\text{Ca}^{2+}$ . The results shown above indicate that the surface membrane in the presence of polyvalent cations behaves in entirely different ways for reception of  $\text{Na}^+$  and  $\text{K}^+$ .

### **Discussion**

The results shown in Fig. 1 indicated that an increase of  $\text{CaCl}_2$  concentration from  $2 \times 10^{-5}$  to  $10^{-4} \text{ M}$  at a given concentration of NaCl was associated with a large decrease in  $\Delta\zeta = \zeta - \zeta_0$  of the slime mold when the concentration of NaCl was higher than the  $C_{th}$  (see chained line *a-b* in Fig. 1). Since the zeta potential is proportional to the surface charge density at the membrane surface at a given ionic strength in medium [10], the decrease in  $\Delta\zeta$  corresponds to an increase in negative charge density at the membrane surface of the slime mold. This increase in the density of negative fixed charge caused by an addition of  $\text{CaCl}_2$  is

not attributable to the  $\text{Ca}^{2+}$  adsorbed on the membrane surface because  $\text{Ca}^{2+}$  has positive charge. Therefore, the increase of the negative charge must be attributed to a conformational change of the surface membrane induced by reception of  $\text{CaCl}_2$ , where either an appearance of negative charge or a disappearance of positive charge is associated. It is interesting to note that the conformational change occurring at the membrane surface by reception of  $\text{CaCl}_2$  could be observed only when the concentration of  $\text{NaCl}$  exceeded the  $C_{th}$ . An increase of the negative surface charge led to an increase of the  $C_{th}$  for  $\text{NaCl}$  in the zeta potential, which in turn led to an increase of the  $C_{th}$  for  $\text{NaCl}$  measured by the membrane potential, because the changes in the zeta potential coincided with those in the membrane potential both in the presence and the absence of  $\text{CaCl}_2$ , as pointed out previously.

The conformational change caused by reception of a chemical stimulus could also be observed when the slime mold was subjected to nonelectrolyte stimuli, such as sugars or alcohols. Although in these cases, the negative charge density at the membrane surface decreased by the reception of nonelectrolytes both for attractants and repellents [10].

The selectivity to univalent cations was affected remarkably by the presence of polyvalent cations. Also, the temperature dependences of the  $C_{th}$  for  $\text{KCl}$  and  $\text{NaCl}$  are different from each other when a polyvalent cation concentration in medium exceeded its own threshold (see Fig. 6). These facts imply that the characteristics of the membrane of the slime mold before and after the reception of a polyvalent cation are quite different from each other. If the concentration of  $\text{Ca}^{2+}$ , for example, in the external solution is decreased successively, the  $C_{th}$  for  $\text{NaCl}$  decreases while that for  $\text{KCl}$  increases discontinuously at  $6 \times 10^{-5} \text{ M}$   $\text{CaCl}_2$  (see Fig. 3). Thus, the selectivity of the membrane to  $\text{K}^+$  and  $\text{Na}^+$  is changed in different directions by the elimination of  $\text{Ca}^{2+}$  from the membrane surface of the slime mold. The different responses of biomembranes between  $\text{K}^+$  and  $\text{Na}^+$  caused by  $\text{Ca}^{2+}$  concentration have been observed in various functions in biological systems as described in the introduction. The data presented here may throw a light on the mechanism of these antagonistic actions between uni- and divalent cations.

The selectivity of univalent cations in biomembranes is one of the central problems in the studies of living cells. Selectivity in the membrane system was analyzed theoretically by Eisenman some years ago [6, 7] (see also [8]) in terms of the hydration energy of cations and the coulomb force acting between ions and the anionic site in the membrane. He predicted that only 11 sequences of cation selectivity are possible in

nature. As shown in the previous paper [23], however, the sequence of cation selectivity observed with the slime mold in pure water was not included in the classifications proposed by Eisenman. The sequence of cation selectivity in the presence of polyvalent cations presented in this paper was also not in line with the Eisenman theory. It would be desirable to investigate more generally the origin of selectivity to cations in membrane systems.

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### References

1. Anderson, J.D. 1964. Regional differences in ion concentration in migrating plasmodia. *In: Primitive Motile Systems in Cell Biology*. D. Allen and N. Kamiya, editors. pp 125–134. Academic Press, New York and London
2. Camp, W.G. 1936. A method of cultivating myxomycete plasmodia. *Bull. Torrey Bot. Club*. **63**:205
3. Carlilie, M.J. 1970. Nutrition and chemotaxis in the myxomycete *Physarum polycephalum*: The effect of carbohydrate on the plasmodium. *J. Gen. Physiol.* **63**:221
4. Coman, D.R. 1940. Additional observation on positive and negative chemotaxis: Experiments with myxomycete. *Arch. Pathol. Lab. Med.* **29**:220
5. Daniel, J.W., Rusch, H.P. 1961. The pure culture of *Physarum polycephalum* on a partial defined soluble medium. *J. Gen. Microbiol.* **25**:47
6. Diamond, J.M., Wright, E.M. 1969. Biomembranes; the physical basis of ion and nonelectrolyte selectivity. *Annu. Rev. Physiol.* **31**:581
7. Eisenman, G. 1962. Cation selective glass electrodes and their mode of operation. *Biophys. J.* **2**:(2):259
8. Eisenman, G. 1969. Theory of membrane electrode potentials: An examination of the parameters determining the selectivity of solid and liquid ion exchangers and of neural ion-sequestering molecules. *In: Ion Selective Electrodes*. R.A. Durst, editor. pp. 1–56. Nat. Bur. Standards, Washington
9. Harris, H. 1961. Chemotaxis. *Exp. Cell Res.* **8** (Suppl.):199
10. Hato, M., Ueda, T., Kurihara, K., Kobatake, Y. 1976. Change in zeta potential and membrane potential of slime mold *Physarum polycephalum* in response to chemical stimuli. *Biochim. Biophys. Acta* **426**:73
11. Inoue, I., Ishida, N., Kobatake, Y. 1973. Studies of excitable membrane formed on the surface of protoplasmic droplets isolated from *Nitella*: IV. Excitability of the drop membrane in various compositions of the external solutions. *Biochim. Biophys. Acta* **330**:27
12. Kamiya, N. 1942. Physical aspects of protoplasmic streaming. *In: The Structure of Protoplasm*. W. Seifritz, editor. pp. 199–244. Iowa State College Press, Ames
13. Kamiya, N. 1959. Protoplasmic Streaming. *Protoplasmatologia* **8**:1
14. Kashiwagura, T., Kamo, N., Kurihara, K., Kobatake, Y. 1977. Enhancement of salt responses in frog gustatory nerve by removal of  $\text{Ca}^{2+}$  from the receptor membrane treated with 1-anilinonaphthalene-8-sulfonate. *J. Membrane Biol.* **35**:205
15. Katsuki, Y., Hashimoto, T., Yanagisawa, K. 1970. The lateral-line organ of shark as a chemoreceptor. *Adv. Biophys.* **1**:1

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16. Kobatake, Y., Inoue, I., Ueda, T. 1975. Physical chemistry of excitable membranes. *Adv. Biophys.* **7**:43
17. Kusano, K. 1958. The influence of cations on the activity of gustatory receptors: II. Effects of NaCl, LiCl, NH<sub>4</sub>Cl and CsCl. *Kumamoto Med. J.* **11**:240
18. Rosen, W.G. 1962. Cellular chemotropism and chemotaxis. *Q. Rev. Biol.* **37**:242
19. Tasaki, I. 1968. Nerve Excitation. C. Thomas, Springfield, Ill.
20. Tasaki, I., Singer, I., Takenaka, T. 1965. Effects of internal and external ionic environment on excitability of squid giant axon. *J. Gen. Physiol.* **48**:1095
21. Terayama, K., Ueda, T., Kurihara, K., Kobatake, Y. 1977. Effect of sugars on salt reception in true slime mold *Physarum polycephalum*: Physicochemical interpretation of interaction between salt and sugar receptions. *J. Membrane Biol.* **34**:369
22. Ueda, T., Muratsugu, M., Kurihara, K., Kobatake, Y. 1976. Chemotaxis in *Physarum polycephalum*: Effects of chemicals on isometric tension of the plasmodial strand in relation to chemotactic movement. *Exp. Cell Res.* **100**:337
23. Ueda, T., Terayama, K., Kurihara, K., Kobatake, Y. 1975. Threshold phenomena in chemoreception and taxis by slime mold *Physarum polycephalum*. *J. Gen. Physiol.* **65**:223
24. Voet, A. 1939. Quantitative lyotropy. *Chem. Rev.* **20**:169